

2 **Screening of endocrine disruption activity in sediments**
3 **from the Uruguay River**

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6 Accepted: 16 April 2014
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8 **Abstract** Sediment constitutes an important sink of
9 endocrine disruptor compounds; however, the potential of
10 sediments to act as a source of endocrine disruptors should
11 be more extensively investigated. The main objective of
12 this study was to determine whether exposure of immature
13 common carp to Uruguay River sediments undergo phys-
14 iological and endocrine alterations. The lower Uruguay
15 River watershed supports intensive agricultural and forest
16 production, receives municipal sewage discharge and
17 industrial effluent, and a new large pulp mill was con-
18 structed in 2006. A 30-day semi-static assay was performed
19 using sediments from four sites along the Uruguay River
20 and compared with an unexposed group in dechlorinated
21 water as a negative control. We focused on two upstream
22 and two downstream sites of a new elemental chlorine free
23 pulp mill. The results showed that plasma vitellogenin
24 levels increased in fish along the river and significant dif-
25 ferences were found between the exposed and unexposed
26 groups. Condition factor and gonadosomatic index were

not different; however, a significant difference in hepato- 27
somatic index was observed in fish exposed to sediment 28
from an industrial site. A significant reduction in primary 29
spermatocyte accumulation was observed in the exposed 30
group compared with that in the control group, and some 31
individuals exposed to sediments from industrial sites 32
presented with testis-ova. Our results suggest that Uruguay 33
River sediments act as an important source of estrogenic 34
compounds that could be responsible for the alterations 35
observed. Future studies are needed to identify the causal 36
agents and determine exposure routes. 38

Keywords Endocrine disruptors · Common carp · 39
Sediment · Vitellogenin · Laboratory-exposure 40

Introduction 41

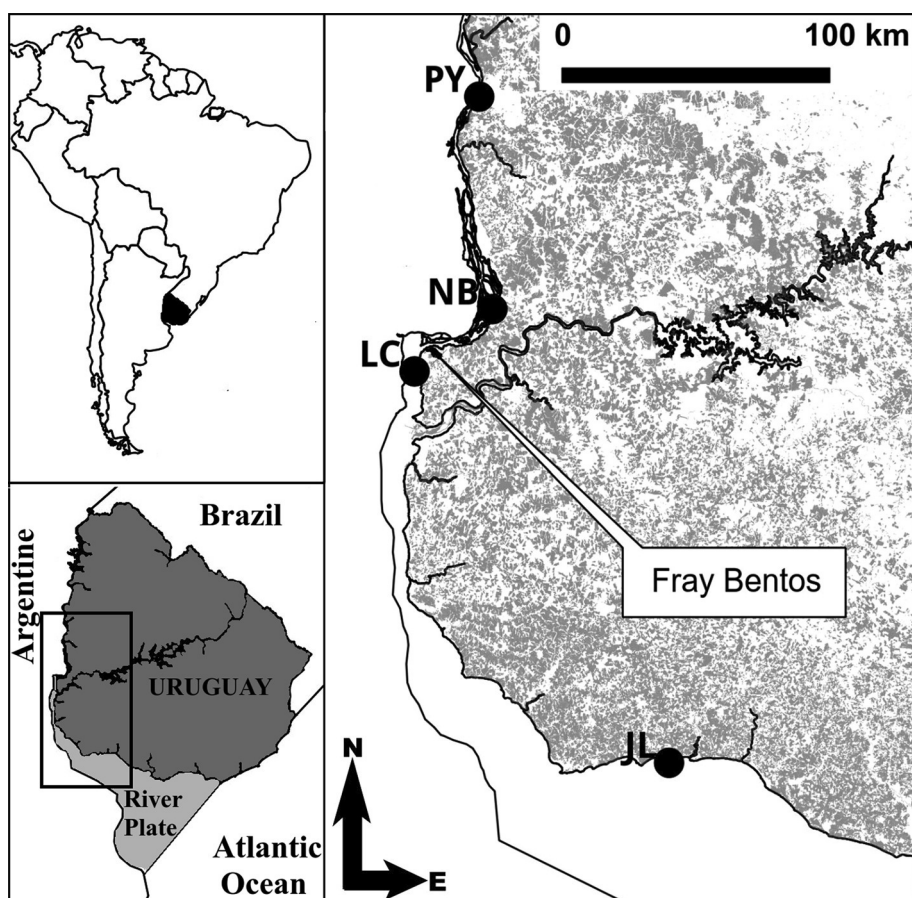
Growing scientific evidence indicates that some natural and 42
synthetic chemical compounds interfere with normal 43
endocrine functioning in humans and wildlife. Most of 44
these studies have focused on detecting specific endocrine 45
disruptor compounds (EDCs) in surface water and their 46
potential effects on fish (Sellin et al. 2009, 2011a; Velisek 47
et al. 2011). Recent studies have detected high concentra- 48
tions of EDCs in sediments, suggesting that the sediments 49
could be responsible for the observed alterations; however, 50
bioavailability of EDCs is complex. Sediments could be 51
acting as a sink and reducing EDC bioavailability or re- 52
releasing the chemical compounds into the water and act- 53
ing as a source. The possible exposure routes to aquatic 54
organisms include direct uptake of free compounds across 55
the gills or skin and ingestion of sediment particles (Peck 56
et al. 2004). Several laboratories and field studies have 57
reported that fish exposed to sediments experience 58

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Fig. 1 Study area and location of the sediment sampling zones. *PY* Paysandú, *NB* Nuevo Berlín, *LC* Las Cañas, *JL* Juan Lacaze. The grey zones on the Uruguayan territory show land under agricultural activity. The information was obtained from supervised classification of LANDSAT 5 satellite images



59 significant alterations in endocrine functions (Orrego et al.
60 2005; Kolok et al. 2007; Sellin et al. 2010, 2011b; Kolpin
61 et al. 2013; Jessick et al. 2014). However, the potential of
62 sediments to act as sources for endocrine disruptors should
63 be more extensively investigated.

64 The lower Uruguay River watershed is characterized by
65 intensive agricultural and forest production. This part of the
66 river receives a variety of municipal sewage discharges and
67 industrial effluents, and a new large pulp mill was con-
68 structed in 2006 (Paruelo et al. 2006; Céspedes-Payret et al.
69 2009; Vega et al. 2009). From 2005 to 2007 several chemi-
70 cals such as resin acids, phytosterols, polychlorinated
71 dibenzo-p-dioxins, and dibenzofurans have been detected in
72 water and sediments from the lower Uruguay River and
73 bioaccumulation and ecotoxicological effects have been
74 observed in wild fish (Míguez et al. 2010; Saizar et al. 2010).

75 This study was designed to test the hypothesis that a
76 variety of chemicals found in sediments can display
77 endocrine disrupting activity. The main goal was to
78 determine if immature common carp (*Cyprinus carpio*)
79 exposed to sediments from the lower Uruguay River
80 exhibit physiological and endocrine alterations as deter-
81 mined by condition factor, hepatosomatic and gonadoso-
82 matic indices, plasma vitellogenin levels, and a gonadal
83 histological analysis.

Materials and methods

Sediment samples

84
85
86 Sediment from four sites along the lower Uruguay River
87 (between S 30°11', W 57°38' and S 34°00', W 58°21')
88 were collected with an Eckman dredge in December 2006,
89 transported at 4 °C to the laboratory, and stored at -20 °C
90 until bioassay. We focused on two upstream sites from a
91 new elemental chlorine free (ECF) pulp mill: the Paysandú
92 (PY) urban-industrial zone and the Nuevo Berlín (NB)
93 agricultural and forested (*Eucalyptus* sp.) zone and two
94 downstream sites: the Las Cañas (LC) agricultural and
95 tourist zone and the Juan Lacaze (JL) urban-industrial
96 zone, which are directly influenced by discharge from an
97 elemental chlorine bleached kraft pulp mill (Fig. 1).

Experimental design

98
99 Immature common carp (mean body length, 7.2 ± 0.9 cm;
100 mean weight, 9.0 ± 3.2 g) were obtained from the
101 National Direction of Aquatic Resources Fish Hatchery.
102 The fish were acclimatized for 15 days in an aerated pool
103 (800 L of dechlorinated water, renewed every 2 days),
104 maintained with a constant temperature (22 ± 1 °C), light:

105 dark cycle (12:12 h), and dissolved oxygen ($89 \pm 1 \%$).
 106 Fish were fed commercial food (Marplatense S.A., Mon-
 107 tevideo, Uruguay) ad libitum.

108 A 30-day semi-static assay (total water renewed every
 109 7 days) was performed in January 2007 using the same
 110 environmental conditions as those used for acclimatization.
 111 Sixty fish were randomly allocated in 30 L aquaria (four
 112 per aquaria) into five groups with 12 fish in each group.
 113 One group was placed in dechlorinated water as a negative
 114 control, and the others were exposed to sediments from
 115 each zone (PY, NB, LC, and JL) a 1:10 w/v rate (Sellin
 116 et al. 2011b; Jessick et al. 2014). No mortalities were
 117 observed during the assay.

118 Plasma vitellogenin (VTG) levels

119 Once the bioassay was completed, blood samples were
 120 extracted from the vena caudalis using heparinized syringes
 121 for plasma VTG analysis. Plasma was separated by
 122 centrifugation (Universal 32R, Hettich Zentrifugen) at
 123 1,500 rpm for 10 min, and plasma VTG was quantified
 124 using antibody pre-coated enzyme-linked immunosorbent
 125 assay kits (Biosense Laboratory, Bergen, Norway; product
 126 no. V01003402) (Nilsen et al. 2004). The microplates were
 127 measured at a wavelength of 492 nm in a Biorad 680
 128 microplate reader spectrophotometer (Hercules, CA, USA).
 129 The VTG concentration was calculated based on a standard
 130 calibration curve and expressed in $\mu\text{g/mL}$.

131 Gonadal histology

132 Gonads were immediately fixed in 10 % phosphate buf-
 133 fered formalin at pH 7.4. They were dehydrated gradually
 134 (alcohol 70, 96, and 100 v/v% chloroform) and embedded
 135 in paraffin. Sections (5 μm , Reichert–Jung microtome)
 136 were rehydrated and stained with hematoxylin and eosin.
 137 The fish were sexed and the reproductive maturity of the
 138 gonad cells was determined according to Smith and Walker
 139 (2004) using an optical microscope (Olympus Vanox;
 140 Tokyo, Japan) and photographed using a digital camera.

141 Physiological indices

142 The hepatosomatic index (HSI) and gonadosomatic index
 143 (GSI) of each fish was determined by dividing the body
 144 mass of the fish by the mass of the tissues and multiplying
 145 by 100. Condition factor (K) was calculated according to
 146 the following equation: $(K) = 100 (\text{body mass without}$
 147 $\text{organs/standard length}^b)$, where b is the allometric coeffi-
 148 cient, estimated by linear regression after logarithmic
 149 transformation of body mass and standard length before
 150 starting the assay ($n = 60$).

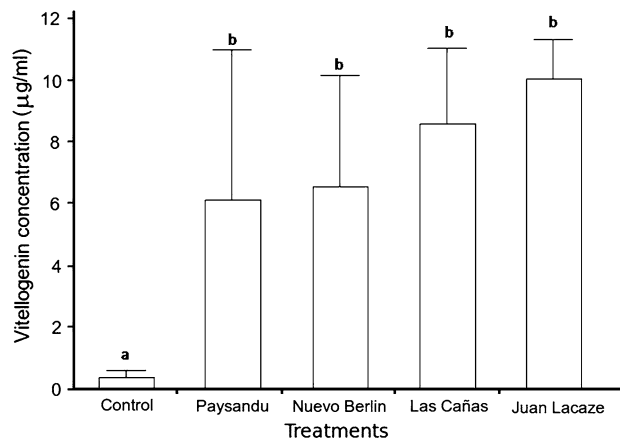


Fig. 2 Plasma vitellogenin levels in common carp exposed to sediments from the Uruguay River. Vertical bars indicate the 95 % confidence intervals. Different letters indicate significant differences

Statistical analysis

Normality and homogeneity of variance were verified and a single factor analysis of variance or Kruskal–Wallis test was used to determine differences between the physiological indices and plasma VTG levels. Statistical significance was confirmed by Tukey's post hoc test A $p < 0.05$ was considered significant.

Results

Plasma VTG levels and gonadal histology

Plasma VTG levels are presented in Fig. 2. The values increased along a latitudinal gradient from PY to JL. Significant differences were observed among the sediment-exposed groups and the control (Tukey's HSD, $p < 0.05$); however, no differences were detected among the exposed groups.

The gonadal histological analysis of females revealed that oocyte stages were not different between exposed and unexposed groups and that all oocytes were in the previtellogenic and perinucleolar stages (Fig. 3C, D). However, sediment-exposed males showed a significant reduction in the number of primary spermatocytes compared with those in the control group ($p = 0.01$) (Fig. 3A, B), and some individuals exposed to sediments from industrial sites (PY and JL) presented with testis–ova (Table 1).

Physiological indices

Mean (\pm standard error) values of the physiological indices in the exposed and unexposed groups are given in Table 2. No significant differences were observed among the groups

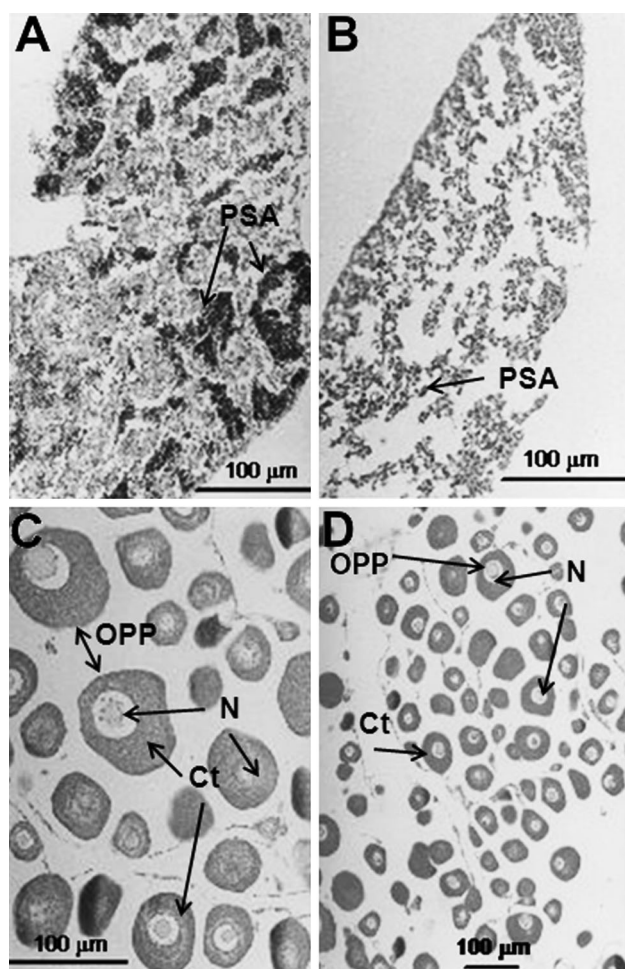


Fig. 3 Photographs of gonad histology in common carp exposed to sediments from different zones along the lower Uruguay River. **a** Gonads of a male from the control (magnification, 20 \times); **b** gonads of a male from the Nuevo Berlín (NB) fish group (magnification, 20 \times); **c** gonads of a female from the control (magnification, 20 \times); **d** gonads of a female from the NB group (magnification, 10 \times). *PSA* primary spermatocyte accumulation, *OPP* oocytes in previtellogenic and perinucleolar stage, *Ct* cytoplasm, *N* nucleus

179 for K or GSI; however, significant differences in HSI were
180 detected. Post-hoc comparisons revealed that fish exposed
181 to sediment from JL had significantly increased HSIs
182 compared with those in the control ($p = 0.04$) and LC
183 groups ($p = 0.02$).

184 Discussion

185 The main goal of this study was to determine whether
186 immature common carp exposed to Uruguay River sedi-
187 ments exhibit physiological and endocrine alterations. Our
188 results clearly indicate that natural and/or synthetic
189 chemical compounds present in the sediments caused an
190 abnormal and significant induction of the egg-yolk

Table 1 Sex distribution between treatments and the control and the occurrence of testis–ova

Treatment	Sex distribution (males/females per treatment)	Testis–ova frequency
Unexposed	9/3	0/9
Paysandú	9/3	1/9
Nuevo Berlín	4/8	0/4
Las Cañas	4/8	0/4
Juan Lacaze	10/2	1/10

Table 2 Physiological indices in the exposed and unexposed groups

Treatment	K $^{\alpha}$	GSI $^{\beta}$	HSI $^{\gamma}$
Unexposed	2.6 \pm 0.4 (12)	0.5 \pm 0.6 (12)	1.3 \pm 0.8 (12) a
Paysandú	2.8 \pm 0.3 (12)	0.5 \pm 0.6 (12)	1.7 \pm 0.6 (12) ac
Nuevo Berlín	2.6 \pm 0.5 (12)	1.3 \pm 1.0 (12)	1.8 \pm 0.6 (12) ac
Las Cañas	2.6 \pm 0.4 (12)	0.9 \pm 0.6 (12)	0.9 \pm 0.6 (12) a
Juan Lacaze	2.8 \pm 0.4 (12)	0.6 \pm 0.8 (12)	2.5 \pm 0.4 (12) bc

Values are given as mean \pm standard deviations for each treatment, and respective n in parentheses

Treatments with statistical differences have different letters

α Condition factor

β Gonadosomatic index

γ Hepatosomatic index

precursor protein VTG in all exposed groups. Elevated
191 levels of VTG in males and immature females were clearly
192 an estrogen-mediated response. The exposure to estrogenic
193 compounds has a transitory effect on VTG production;
194 however, exposure during early life stages could reduce
195 survivability, and the effects on gonadal morphology and
196 reproductive function may be permanent (Hutchinson et al.
197 2006).
198

Several known sources of endocrine disruptors are
199 located in the Uruguay River watershed, and previous
200 studies have detected some EDCs in sediments (Saizar
201 et al. 2010) that could be responsible for the observed
202 alterations. The urban-industrial sites (PY and JL) receive
203 untreated municipal sewage effluent containing a complex
204 cocktail of natural (estrone or 17 β -estradiol) and synthetic
205 estrogens used in oral contraceptives as well as surfactants
206 used in soaps and detergents (alkylphenols and alkylphe-
207 nolphenoxyethoxylates). Furthermore, the plasma VTG con-
208 centrations in fish were highest where deposition processes
209 were predominant at JL and where pulp mill effluent was
210 discharged near the sampling site.
211

The agricultural and forest river sectors (NB and LC)
212 support intense soybean-wheat row crop production, and
213 this agricultural system uses known estrogenic pesticides
214

215 such as chlorpyrifos, endosulfan, and cypermethrin (Mnif
216 et al. 2011). Additionally, phytoestrogens released by crops
217 as a defense strategy may be reaching the river in overland
218 runoff. In particular, soybeans contain high levels of gen-
219 istein, daidzein, and glycitein, which can elicit alterations
220 in endocrine function in wildlife and humans (Ng et al.
221 2006). It is important to note that VTG levels were not
222 affected by the sex ratio, as shown by similar VTG con-
223 centrations in the LC and JL groups with opposite sex
224 ratios. This was also true when comparing the PY and NB
225 treatments.

226 The significant increase in liver mass at JL may have
227 been caused by induction of the hepatic mixed function
228 oxidase system in response to discharge of persistent
229 organic compounds from the pulp mill effluent. Increased
230 protein synthesis generates proliferation of endoplasmic
231 reticulum, which can be reflected in increased hepatocyte
232 size (Munkittrick et al. 1992).

233 The gonad histology analyses indicated that female fish
234 did not exhibit differences in maturation state; however,
235 sediment-exposed males presented delayed testicular ma-
236 turation than that in the unexposed group. Jobling et al.
237 (1996) reported that the induction of VTG in males is
238 negatively correlated with testicular maturation, and Dev-
239 lin and Nagahama (2002) observed retarded gonadal ma-
240 turation in *C. carpio* males exposed to estrogenic
241 compounds. Changes in sex ratios and intersex individuals
242 have been reported in common carp exposed to EDCs
243 (Gimeno et al. 1998; Devlin and Nagahama 2002). How-
244 ever, the intersex condition occurs naturally in approxi-
245 mately 5 % of the population in this species (Jobling et al.
246 1998). Thus, the presence of individuals with testis-ova
247 observed in our study was possibly a natural phenomenon
248 and may not have been caused by exposure to contami-
249 nated sediments.

250 Conclusions

251 This study is the first report about endocrine disruption in
252 fish exposed to sediment from the lower Uruguay River.
253 The results can be considered a reference condition for
254 monitoring the impacts of the new ECF bleached kraft
255 *Eucalyptus* pulp mill. Nonpoint (soybean-wheat crops) and
256 point sources (municipal sewage and pulp mill effluent)
257 can explain the VTG induction observed in immature fish,
258 and suggest the presence and bioavailability of EDCs in the
259 sediments. The specific agents responsible for the toxic
260 effects were not identified because it was beyond the scope
261 of this study. Future research is needed to identify the
262 causal agents (natural or synthetic) and to determine
263 exposure routes (e.g., grazing on sediments or bioconcent-
264 ration from the water column).

Acknowledgments We express our thanks to the Departments of
Virology and Cellular Biology (Science School) for cooperation with
the processing of samples and to Mr. Angel Rosano and the Ur-
guayan Navy for their assistance in the fieldwork. This study was
funded by the National Agricultural Research Institute (INIA) Project
SA07 and the Environmental Science Master Program.

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